

1 **Rightsizing root phenotypes for drought resistance**

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9 **Highlight:** Parsimonious root phenotypes may benefit water capture under drought.

10

11 **Abstract**

12 I propose that reduced root development would be advantageous for drought resistance
13 in high-input agroecosystems. Selection regimes for crop ancestors and landraces
14 include multiple stresses, intense competition, and variable resource distribution, which
15 favored prolific root production, developmental plasticity in response to resource
16 availability, and maintenance of unspecialized root tissues. High-input agroecosystems
17 have removed many of these constraints to root function. Therefore, root phenotypes that
18 focus on water capture at the expense of ancestral adaptations would be better suited to
19 high-input agroecosystems. Parsimonious architectural phenotypes include fewer axial
20 roots, reduced density of lateral roots, reduced growth responsiveness to local resource
21 availability, and greater loss of roots that do not contribute to water capture. Parsimonious
22 anatomical phenotypes include reduced number of cortical cell files, greater loss of
23 cortical parenchyma to aerenchyma and senescence, and larger cortical cell size.
24 Parsimonious root phenotypes may be less useful in low-input agroecosystems, which
25 are characterized by multiple challenges and tradeoffs for root function in addition to water
26 capture. Analysis of the fitness landscape of root phenotypes is a complex challenge that
27 will be aided by the development of robust functional-structural models capable of
28 simulating the dynamics of root-soil interactions.

29 Keywords: root development; drought; root architecture; root depth; root anatomy,
30 phosphorus, nitrogen, water

31 **Introduction**

32 Here I consider the hypothesis that root phenotypes for optimal crop yield under drought
33 differ from those resulting from natural selection, domestication, and traditional crop
34 breeding. Specifically, I consider the hypothesis that reduced root development has
35 benefits for crop yield under water stress. I focus on annual crops grown for seed yield,
36 but many elements of this consideration pertain to annual crops grown for biomass yield
37 and perennial crops. Some elements of my discussion are supported by the research
38 literature, others are more speculative. I will not attempt a comprehensive review of all
39 relevant literature but where appropriate cite recent reviews.

40 In the large majority of agroecosystems, seeds are planted in moist soil, so that soil
41 moisture is adequate for germination and emergence. As soil moisture is depleted the
42 crop becomes increasingly reliant on precipitation to meet its water requirement. A
43 common scenario is progressively increasing water deficit over time as soil moisture is
44 exhausted, crop growth increases water demand, and evapotranspiration increases as
45 the season progresses. Sporadic rainfall may be inadequate to meet evapotranspiration,
46 causing progressively increasing water stress over time, the extreme scenario being
47 terminal drought. In other cases, temporary water deficit develops earlier in the season,
48 which can have severe effects on reproduction by affecting seed formation or fertilization.
49 I focus here on progressive water stress as the archetypal drought scenario, in which
50 water is more available in deep soil strata than at the soil surface over time. Some
51 elements of my discussion also pertain to sporadic drought scenarios in which water is
52 available in surface strata. I also focus on growth itself rather than specific effects of water
53 stress on reproduction, which are important (Saini and Westgate, 2000), and I focus on
54 phenes states ('phene' is to 'phenotype' as 'gene' is to 'genotype', (York *et al.*, 2013)) that
55 do not incur inherent yield penalties (Blum, 2005), as is the case, for example, with shorter
56 phenology (Nord and Lynch, 2009).

57 **1. Do crop plants make too many roots?**

58 Terrestrial plants evolved in environments that favor phenotypes with aggressive root
59 proliferation. The soil is a hostile environment for roots. Soils can be too dry or too wet

60 (hypoxic) to sustain root tissue, can be too hard to permit root penetration, can present
61 toxins or deficiencies that directly limit root growth (e.g. Ca deficiency, Al toxicity), and are
62 inhabited by many herbivores and pathogens, resulting in substantial root turnover (Fisher
63 *et al.*, 2002). In natural ecosystems, intense belowground competition means that failure
64 to produce sufficient roots to capture soil resources will cede a competitive advantage to
65 neighbors. Furthermore, the availability of soil resources in time and space is nonuniform
66 and in many cases unpredictable. The three primary soil resources that limit plant growth
67 in the vast majority of terrestrial ecosystems are N, P, and water. In those rare soil
68 environments that have abundant N, P, and water, dense plant growth results in intense
69 belowground competition, which reduces the availability of these resources for individual
70 plants. Phosphorus is generally more available in surface horizons because of continual
71 deposition on the soil surface in plant residues, and its low mobility in soil (Lynch, 2011).
72 However shallow soil strata may become hostile to root foraging because of temperature
73 extremes (both hot and cold) as well as drying from root uptake and direct evaporation.
74 Nitrogen in natural systems is released from mineralization of soil organic matter through
75 microbial activity, forming ammonium, which is slowly mobile, which is in turn readily
76 converted to nitrate, which is highly soluble in water and therefore can move to deeper
77 soil strata through leaching. Since soil organic matter is concentrated in shallow soil
78 strata, its mineralization is subject to variation in temperature and water availability. The
79 leaching of nitrate to deeper soil strata is also subject to water availability and soil physical
80 conditions. Therefore, the spatiotemporal availability of N, like P, is subject to water
81 availability (Thorup-Kristensen and Kirkegaard 2016; Dathe *et al.*, 2016). Since water
82 availability (generally in the form of precipitation) is itself highly stochastic in time and
83 space, N and P availability in time and space are also to some extent unpredictable.
84 Because of the unpredictability of resource availability, belowground competition, and the
85 inevitability of root loss, plants have been selected for aggressive root proliferation and
86 architectural plasticity, resulting in focused proliferation in response to patches of the
87 primary resources of N, P, and water.

88 The strategy of aggressive root production may be counterproductive in high-input
89 agroecosystems. In such systems, intensive fertilization has removed P and N as growth
90 limitations. Indeed, overuse of fertilizers in such systems has created substantial

91 environmental pollution with global repercussions. Intensive application of pesticides,
92 modern crop management, and ongoing crop breeding for pest resistance have
93 significantly reduced the risk of root loss from pathogens, herbivores, and some abiotic
94 stresses that are important in natural systems such as soil acidity. Herbicides and
95 reduced tillage, coupled with herbicide-resistant GM crops have substantially reduced
96 root competition with weeds. Many of the technological elements of modern crop
97 production can be viewed as means to eliminate constraints to root function. High-input
98 agriculture has therefore mitigated many of the constraints to root function that are
99 important in the ecosystems in which crop ancestors evolved, and in which crops were
100 domesticated. As a result, we cannot assume that the ancestral strategy of producing
101 many roots and aggressively exploiting resource patches is adaptive in high-input
102 agroecosystems.

103 The one stress factor that has remained important throughout plant evolution and crop
104 domestication, and that remains important in modern agroecosystems, is drought. It is
105 probable that drought stress will increase in the future as a result of global climate change,
106 which will increase atmospheric temperature and thereby soil water loss and crop water
107 demand, and is projected to alter precipitation patterns in time and space, which can have
108 large effects on crop water demand and water availability even if total annual precipitation
109 is unchanged (Nord and Lynch, 2009). The increasing probability of drought is coinciding
110 with both substantially increasing demand for agricultural production from a growing
111 human population as well as increasing degradation of fresh water resources. 'High input'
112 therefore does not include irrigation in most regions. There are insufficient fresh water
113 resources to meet global demand for agricultural production, and the gap between fresh
114 water resources and crop water demand is projected to grow. The importance of drought
115 for high-input agroecosystems is therefore a paramount challenge of the 21st century.
116 Drought stress has always been a primary factor in the evolution of terrestrial plants, but
117 in high-input agriculture it has become the single greatest production risk.

118 The ancestral root phenotype of aggressive root proliferation results from selection
119 regimes characterized by multiple biotic and abiotic constraints. It has been proposed that
120 this phenotype is actually maladaptive for drought resistance (Lynch, 2013). Specifically,

121 it has been proposed that root phenotypes for superior drought resistance should have
122 fewer axial roots, fewer lateral roots, and less architectural plasticity in response to
123 resource patches ((Lynch, 2013), Fig. 1). Recent evidence supports elements of this
124 hypothesis, as summarized below. The idea that parsimonious root phenotypes in general
125 are useful for drought resistance is supported by van Oosterom et al (2016), who showed
126 that drought resistance in maize is associated with 'root system efficiency' (RSE), defined
127 as transpiration per unit leaf area per unit of root mass, meaning that genotypes capable
128 of maintaining transpiration at low root mass are advantageous. Another study with maize
129 showed that lines resulting from 8 cycles of selection for grain yield under drought had
130 33% less root biomass in the topmost 50 cm of soil (Bolaños *et al.*, 1993). These studies
131 support the concept that more parsimonious root phenotypes are useful under drought,
132 but are not mechanistic, since root biomass is a coarse metric aggregating many distinct
133 root phenes and phene states. Evidence for the utility of parsimonious root phenes for
134 drought resistance is summarized below.

135 **1a. Evidence that reducing the number of axial roots improves drought resistance**

136 A recent study supports the hypothesis that reduced production of axial roots is
137 advantageous under drought (Gao and Lynch, 2016). Maize recombinant inbred lines
138 with shared genetic backgrounds but with contrasting production of crown roots (*i.e.*
139 belowground nodal roots, which are the dominant class of axial roots in this species) were
140 grown under water deficit stress in field rainout shelters and soil mesocosms. Water
141 stress reduced the production of crown roots, and lines with fewer crown roots under
142 stress had substantially deeper rooting and greater capture of subsoil water, and
143 consequently improved plant water status, stomatal conductance, leaf and canopy
144 photosynthesis, biomass, and yield (Fig. 2).

145 The Gao and Lynch (2016) study observed that water stress reduced the production of
146 axial roots in maize. While growth reduction under stress could simply be a symptom of
147 stress injury, it could also reveal an adaptation mechanism, especially if it is regulated by
148 stress response pathways in a coordinated manner. In this context, it is noteworthy that
149 water stress indeed downregulates the production of crown roots in grasses through

150 coordinated developmental pathways (Sebastian *et al.*, 2016). Sebastian *et al.* found that
151 water deficit inhibits the production of crown roots in several grass species including
152 *Setaria viridis*, *Setaria italica*, and both wild (teosinte) and cultivated *Zea mays*, by local
153 sensing of soil water availability. Water deficit inhibited crown root production more
154 strongly in wild taxa than in cultivated taxa, and genetic variation was observed in crown
155 root arrest among maize lines. Under water deficit a maize mutant that lacks the ability to
156 form nodal roots (*rtcs*) maintained better soil and plant water status (but did not have
157 different shoot biomass) than a wildtype line. The authors propose that "... severe
158 reductions in shoot-borne root growth are crucial to prevent overdraw of water from
159 the soil and water loss through crown root growth." (Sebastian *et al.*, 2016). This is an
160 important study that generally supports the hypothesis that phenotypes with fewer nodal
161 roots are useful under water stress. However, the ecophysiological interpretation of these
162 results is debatable. It is unclear why conserving soil moisture under water deficit would
163 be advantageous to wild plants subject to competition. Water left in the soil by reduced
164 root growth would be subject to loss to competitors. We might assume that when
165 competition is limited to monogenetic stands as occurs in high-input agriculture, a 'water
166 banking' strategy may be useful for stand-level reproductive output. The observation that
167 cultivated taxa have less rather than more root inhibition by water stress argues against
168 this however. The proposal that roots directly consume significant amounts of water is
169 unsubstantiated, since the amount of water stored in root tissue is only a very small
170 fraction of the amount of water transpired. A gram of dry root tissue might require 1.5 g
171 of water for hydration (Guo *et al.*, 2013) yet a gram of dry plant tissue requires 250-350 g
172 of transpired water in C4 plants and 650-800 g of transpired water in C3 plants (Ehleringer
173 and Monson, 1993). An alternate ecophysiological interpretation is that reduced
174 production of nodal roots reduces competition among root axes of the same plant for both
175 internal resources (photosynthates) and external resources (in this case, water), meaning
176 that a phenotype with fewer axial roots may actually have greater root depth and greater
177 water acquisition from deep soil strata (Fig. 3, Saengwilai *et al.*, 2014b; Gao and Lynch,
178 2016). This is an important conceptual difference since root phenotypes that reduce
179 whole-plant water acquisition are likely to also reduce yield, because of the close
180 association of shoot water use with shoot C gain (Blum, 2009). While root phenotypes

181 that conserve soil water are useful in systems that rely on stored soil water, their yield
182 penalty reduces their utility in rainfed systems with variable water stress. The
183 ecophysiological interpretation of Sebastian *et al.* is based on comparison of one wildtype
184 phenotype with one mutant with a severely reduced root phenotype grown as young
185 seedlings in small containers of synthetic media in growth chambers at unspecified light
186 intensity. No growth benefit associated with root phenotypes was observed. Greater root
187 depth improves soil water capture in the field and in large mesocosms but may not do so
188 in small containers with limited water supply. A more rigorous ecophysiological analysis
189 would compare stands of mature plants with a range of varying crown root production in
190 actual soil with normal transpiration regimes. This was the approach employed by Gao
191 and Lynch, who observed that under water stress, phenotypes with reduced crown root
192 production had greater acquisition of subsoil water rather than less, which contradicts the
193 'water banking' hypothesis (Gao and Lynch, 2016).

194 Indirect evidence for reduced intraplant competition as the physiological benefit of
195 reduced nodal root production under drought comes from a study of the effect of genotypic
196 variation in crown root production in maize under suboptimal N availability (Saengwilai *et*
197 *al.*, 2014b). Under N limitation, N can be localized in deeper soil strata over time because
198 of leaching, which is comparable to the deep localization of water under progressive
199 drought. In this study maize lines with fewer crown roots had deeper rooting, better
200 capture of subsoil N, and consequently better growth and yield under N stress than
201 genetically related lines with more crown roots (Saengwilai *et al.*, 2014b). The fact that
202 reduced production of nodal roots increases N acquisition from deep soil strata, which is
203 similar to the enhanced capture of deep soil water by maize lines with fewer crown roots
204 (Gao and Lynch, 2016), argues against the 'resource banking' hypothesis. It is noteworthy
205 that N stress, like water stress, reduced crown root production in some maize lines, which
206 may suggest that these stresses share signaling pathways in regulating the production of
207 nodal roots.

208 An additional indirect line of evidence for the utility of reduced crown root production is
209 the observation that the past 100 years of maize breeding has resulted in root phenotypes
210 with fewer nodal roots (Fig. 4, York *et al.*, 2015). During this period maize planting density

211 has more than doubled and yield has increased by a factor of 8, resulting in greater
212 belowground competition and resource efficiency. Inadvertent selection for reduced
213 production of nodal roots would be expected if this phenotype was useful for more efficient
214 capture of soil resources. Indeed, disaggregation of maize root phenes using the
215 functional-structural model *SimRoot* showed that the observed reduction in nodal root
216 production makes the root phenotypes of lines from a century ago as productive and fit
217 as modern root phenotypes in modern production environments (Fig. 5, York *et al.*, 2015).

218 The production of axial roots in tillering species, an important group of grain crops that
219 includes wheat, rice, barley, oat, rye, and millet is complicated by the fact that the majority
220 of nodal roots in these species are produced by tillers. Reduced tillering can be a useful
221 water banking strategy in semi-arid and Mediterranean environments with predictable
222 water limitation (Blum, 2005), but reduced tillering also reduces yield potential, which
223 limits its utility in normal rainfed systems. In rice, reduced nodal root number is associated
224 with improved drought resistance (Catolos et al 2017). The utility of reduced nodal root
225 production per tiller merits attention.

226 **1b. Evidence that reducing the density of lateral roots improves drought resistance**

227 Reduced production of lateral roots and the associated phenotype of longer lateral roots
228 has been proposed to be beneficial under drought (Lynch, 2013, Fig. 3). The rationale for
229 this proposal is the same as that described in section (1a) for reduced production of axial
230 roots, *i.e.* that for a mobile soil resource like water, the production of too many lateral
231 roots is counterproductive by increasing intraplant competition for internal resources
232 (primarily carbohydrates) needed for root growth, as well as competition for the capture
233 of mobile soil resources, in this case water.

234 Direct evidence in support of this hypothesis was provided by analysis of maize
235 recombinant inbred lines sharing genetic backgrounds but with contrasting production of
236 lateral roots grown under water stress in field rainout shelters, in natural drought
237 conditions in the field, and in soil mesocosms (Zhan *et al.*, 2015). Water stress reduced
238 lateral branching of crown roots, and lines with fewer lateral roots under stress had
239 substantially deeper rooting, greater capture of subsoil water, and consequently improved

240 plant water status, stomatal conductance, leaf photosynthesis, biomass, and yield (Fig.
241 6).

242 Indirect evidence supporting the utility of this phenotype is provided by the 'era' study of
243 root phenotypes from 100 years of maize breeding (York *et al.*, 2015). This study found
244 that modern maize root phenotypes have greater distance from the point of crown root
245 emergence from the shoot to the first lateral root, a phenotypic state that effectively reduces
246 lateral root production at the organismic level, as well as longer lateral roots.

247 **1c. Is architectural plasticity useful under drought?**

248 It has been proposed that unresponsiveness of lateral branching to localized resource
249 availability would be advantageous under drought (Lynch, 2013). Localized root
250 proliferation in response to local water availability may be counterproductive because
251 water is an ephemeral resource subject to movement and depletion, whereas root growth
252 is relatively slow, and building and maintaining roots incur significant long-term costs,
253 especially considering that roots are not actively senesced (Fisher *et al.*, 2002). In
254 addition, construction of roots in moist soil domains may incur opportunity costs of failing
255 to build roots in domains that may have greater water availability over time. For example,
256 intermittent rain under drought may cause shallow wetting of surface soil that is rapidly
257 depleted. Root production in shallow soil in response to this ephemeral resource would
258 divert resources from exploitation of deeper strata with greater water availability (Lynch,
259 2013). In natural ecosystems and low-input agroecosystems, exploitation of localized
260 water patches through root plasticity can confer a competitive advantage, but in high-
261 input monoculture this is less important. Indirect evidence for this is the observation by
262 that axial root development in cultivated taxa is less sensitive to local water status than in
263 wild taxa (Sebastian *et al.*, 2016). The utility of root plasticity under drought is unclear and
264 merits research.

265 **1d. Can root lifespan be optimized under drought?**

266 Substantial root loss occurs because of both abiotic and biotic stress. Root loss due to
267 biotic stress (pathogens, root-feeding insects, nematodes, etc.) is mitigated by pesticides

268 in high-input systems, but drought stress increases root loss (Huck *et al.*, 1987). The
269 functional impacts of root loss have been analyzed via microeconomic analysis, in which
270 costs and benefits are compared (Eissenstat and Yanai, 1997). The cost of root loss for
271 plant function includes 1) the loss of the nonrecoverable resources in the lost tissue (*e.g.*
272 energy and material invested in cell wall construction); 2) the opportunity cost of reduced
273 soil resource capture, anchorage, lateral root formation, protective barriers, etc. from the
274 lost segment; and 3) loss of these resources and functions from roots subtending the lost
275 segment. Benefits resulting from root loss include termination of the metabolic costs of
276 maintaining the lost segment, primarily carbohydrate lost to respiration, and allocation of
277 primary resources such as N and P invested in root tissue, and the opportunity to allocate
278 future resources to more productive soil domains. The C costs of maintaining root tissue
279 exceed the C costs of constructing root tissue after a short initial period (Lambers *et al.*,
280 1996), and are a significant component of daily C budgets, especially under edaphic
281 stress (*e.g.*, Nielsen *et al.*, 1998). It is therefore possible that root loss may be beneficial,
282 if the benefit of reduced maintenance costs exceeds the cost of the lost root functions
283 (Eissenstat and Yanai, 1997; Steingrobe *et al.*, 2001). Under drought stress, the soil
284 typically dries from the top down because of greater root activity in the topsoil as well as
285 direct evaporation of soil water. The death of roots in dry topsoil may be beneficial by
286 reducing the cost of maintaining roots that are not contributing to water capture. This is
287 especially true for fine roots, since they are more metabolically active per unit root mass
288 and can be replaced after soil rewetting by lateral branching of axial roots. The loss of
289 shallow roots would permit greater resource allocation to deep roots, which would
290 improve water capture. Compensatory regrowth following mechanical root loss has been
291 observed in common bean (Rubio and Lynch, 2007). It has been proposed that root
292 turnover can increase the capture of the immobile nutrients P and K by permitting greater
293 exploration of new soil, although evidence for this comes from simulation models that do
294 not simulate intraplant competition in a robust manner, and from empirical studies with
295 ingrowth cores that restrict competition and soil resource depletion (Yanai *et al.*, 1995;
296 Steingrobe *et al.*, 2001). The proposal that root loss may be beneficial under drought is
297 speculative and is difficult to verify, since the rate, timing, and position of root loss are
298 difficult to control in empirical studies. *In silico* approaches using modern functional-

299 structural models that explicitly model the spatiotemporal dynamics of root growth and
300 soil resource capture would be useful in this context.

301 **2. Do crop roots make too much tissue?**

302 Section 1 considered root phenotypes that optimize water capture under drought by
303 reducing investment in organs with an unfavorable cost/benefit ratio. This section
304 considers optimization of root phenotypes for water capture by reducing investment in
305 cells and tissues with an unfavorable cost/benefit ratio.

306 **2a. Maize genotypes with fewer root cortical cell files have superior drought** 307 **resistance**

308 A large portion of primary root structure is cortical parenchyma that is metabolically active
309 but has no specialized function in water acquisition. Cortical parenchyma is important in
310 radial transport of water from the epidermis to the stele however. In older root segments
311 this function might be reduced without compromising drought resistance, because older
312 root segments are generally located in soil domains in which water has already been
313 depleted, and older root segments can serve axial transport, relying on subtending
314 segments, including lateral branches, for resource capture. In monocotyledonous crop
315 species that lack secondary growth, the cortex can remain viable for extended periods. It
316 was proposed that maize lines with reduced cortical tissue would have reduced metabolic
317 costs of soil exploration, which would enable greater rooting depth and water capture
318 under drought (Lynch, 2013; Lynch *et al.*, 2014). In a study testing this hypothesis, maize
319 lines with contrasting cortical tissue expressed as *Living Cortical Area* (LCA: total
320 transversal root cortical area minus Root Cortical Aerenchyma (RCA) area and
321 intercellular air space area) were exposed to drought stress in soil mesocosms (Jaramillo
322 *et al.*, 2013). Lines with less LCA had less respiration, greater elongation of axial roots,
323 and better growth under water stress than lines with more LCA. The primary components
324 of LCA are the number of cortical cell files (CCFN, Fig. 7b) and cortical cell size (CCS,
325 Fig. 7c), minus intercellular air space and the tissue lost to aerenchyma formation (RCA,
326 Fig. 7a). It was therefore hypothesized that reduced CCFN, CCS, and RCA could all

327 contribute to water capture under drought through their effects on the metabolic cost of
328 soil exploration (Lynch, 2009).

329 The hypothesis that reduced CCFN improves water capture under drought was tested by
330 comparison of maize lines with contrasting CCFN under water stress in soil mesocosms
331 and field environments in the USA and Malawi. Lines with fewer cortical cell files had less
332 respiration, greater root depth, greater exploitation of deep soil water, and therefore better
333 water status, growth, and substantially greater yield than comparable lines with greater
334 more cortical cell files (Fig. 7, Chimungu *et al.*, 2014b). Substantial genetic variation for
335 CCFN is present in maize (Burton *et al.*, 2013; Chimungu *et al.*, 2014b), so this phe
336 merits attention as a potential breeding target for improved drought resistance.

337 **2b. Loss of root cortical tissue via aerenchyma formation improves drought**
338 **resistance**

339 Many crop species form aerenchyma in the root cortex through programmed cell death.
340 The conversion of living cortical cells to air space allows reinvestment of nutrient
341 resources to other plant tissues, including growing root tips, and reduces the metabolic
342 cost of maintaining the root cortex. It was therefore hypothesized that RCA formation is
343 advantageous for soil exploration and the capture of soil resources under edaphic stress
344 (Lynch and Brown, 1998; Fan *et al.*, 2003). *In silico* analysis in *SimRoot* indicates that the
345 effects of RCA formation on root metabolic costs can improve soil exploration and the
346 capture of N, P, and K when those resources limit growth (Postma and Lynch, 2011).
347 Empirical support for improved N capture with greater RCA formation was provided by
348 comparison of maize lines with contrasting RCA formation under suboptimal N regimes
349 in the field in Africa and North America as well as in soil mesocosms (Saengwilai *et al.*,
350 2014a). Under N stress, high RCA genotypes had less root respiration and N content,
351 and greater rooting depth, N capture, growth and yield. The benefit of RCA for N capture
352 suggests that it may also be useful for water capture, as both nitrate and water tend to be
353 deep soil resources (Thorup-Kristensen and Kirkegaard 2016; Dathe *et al.*, 2016). It was
354 therefore proposed that RCA formation would improve water capture under drought
355 (Lynch, 2013). Under water stress in field rainout shelters and soil mesocosms, maize

356 genotypes with greater RCA formation had reduced root respiration and greater rooting
357 depth, water capture, growth, and yield than related lines with less RCA (Zhu *et al.*, 2010)
358 These results were recently supported by an on-farm analysis of maize lines grown under
359 natural drought conditions by smallholder farmers in Malawi, which showed that lines with
360 greater RCA formation had better water status, growth, and yield than lines with less RCA
361 (Fig. 8, Chimungu *et al.*, 2015b).

362 It has been proposed that RCA may be beneficial under drought by reducing radial water
363 conductance, thereby slowing the depletion of soil water (Vadez, 2014). In addition to
364 conserving soil water for later growth, limited water uptake under drought is also likely to
365 incur benefits from reduced shoot growth hence reduced water demand, more efficient
366 use of water in leaf transpiration, and maintenance of wetter, softer soil surrounding
367 growing root tips (Lynch *et al.*, 2014). RCA formation does reduce the radial transport of
368 water (Fan *et al.*, 2007) and nutrients (Hu *et al.*, 2014). While reduced radial water
369 transport may be a benefit of RCA under drought, several lines of evidence suggest that
370 the effects of RCA on the metabolic costs of soil exploration are important. One line of
371 evidence is *SimRoot* modeling, which correctly predicts benefits of RCA for nutrient
372 capture based on reduced metabolic costs without considering reduced water transport.
373 Another line of evidence is that root phenes apart from RCA that reduce root metabolic
374 costs also improve drought resistance despite the fact that their effects on radial water
375 transport are unclear, including reduced production of axial roots (Gao and Lynch, 2016),
376 reduced lateral root branching (Zhan *et al.*, 2015), reduced CCFN (Chimungu *et al.*,
377 2014b), and increased CCS (Chimungu *et al.*, 2014a). It is also noteworthy that lateral
378 roots have less RCA than axial roots, and lateral roots are more important for soil
379 exploration and water capture. It would be challenging to directly assess the relative
380 importance of these two effects of RCA, *i.e.* effects on radial water transport *vs.* effects
381 on the metabolic costs of soil exploration. These effects could be uncoupled *in silico*, as
382 was accomplished for example to uncouple the effects of RCA formation on reduced C
383 costs *vs.* nutrient remobilization in *SimRoot* (Postma and Lynch, 2011). For this to occur
384 we need more robust models of dynamic interactions of root phenotypes with soil water
385 availability.

386 Substantial variation for RCA among crop genotypes, and its apparent utility under
387 drought, nutrient stress, and hypoxia suggest that it merits attention as a potential
388 breeding target (Lynch *et al.*, 2014; Lynch and Wojciechowski, 2015).

389 **2c. Does loss of root cortical tissue via cortical senescence improve drought**
390 **resistance?**

391 Root cortical senescence (RCS) occurs in several important species including wheat
392 (*Triticum aestivum*), triticale (*Triticosecale*), barley (*Hordeum vulgare*, Yeates and Parker,
393 1985; Liljeroth, 1995), rye (*Secale cereale*) (Deacon and Mitchell, 1985; Jupp and
394 Newman, 1987) and oat (*Avena sativa*) (Yeates and Parker, 1985). Unlike RCA formation,
395 which typically leaves files of living cortical cells connecting the epidermis and
396 endodermis, RCS results in complete loss of living cortical tissue. By analogy with RCA,
397 RCS may be beneficial under drought by reducing the metabolic costs of soil exploration
398 and hence water capture. In barley, RCS substantially reduces root respiration and
399 nutrient content, as well as radial transport of water, nitrate, and phosphate (Schneider *et*
400 *al.*, 2017b). RCS was increased by N and P limitation, and a landrace had greater RCS
401 than a modern cultivar (Schneider *et al.*, 2017b), which is indirect evidence that it may be
402 adaptive under edaphic stress. Functional-structural modeling in *SimRoot* showed that
403 RCS could substantially improve the growth of barley under suboptimal availability of N,
404 P, or K, with the main benefit due to nutrient remobilization from senesced tissue
405 (Schneider *et al.*, 2017a). The ability of RCS to reduce root metabolic costs and thereby
406 to increase rooting depth, and to reduce the radial hydraulic conductance of older root
407 segments suggests that it may be useful for water capture from drying soils (Schneider
408 and Lynch in press, Fig. 9).

409 **2d. Maize genotypes with larger root cortical cells have superior drought resistance**

410 Greater cortical cell size (CCS) could reduce the metabolic cost of soil exploration, and
411 thereby water capture under drought, since larger cells have proportionately more volume
412 occupied by the vacuole, which has less N, P and respiratory cost than does cytoplasm.
413 Direct evidence to support this proposal is provided by a comparison of maize lines with
414 contrasting CCS grown under drought in soil mesocosms, field rainout shelters in the

415 USA, and natural drought in the field in Malawi (Chimungu *et al.*, 2014a). Maize lines with
416 greater CCS had less root respiration, greater root depth, and therefore greater capture
417 of subsoil water, better water status, photosynthesis, growth, and yield (Fig. 7).

418 These results are consistent with the hypothesis that reducing root metabolic costs
419 increases water capture from drying soil. Reduced production of axial roots and lateral
420 roots allow remaining roots to reach deep soil water. Reduced cortical burden, by
421 increased aerenchyma formation, decreased number of cortical cell files, or increased
422 cell size, increases water capture under drought by increasing internal resources
423 available for deeper rooting. Root cortical senescence and some degree of root loss may
424 have similar benefits.

425 **2e. Overcoming soil impedance is an important feature of drought resistance**

426 In most soils, soil impedance increases with soil depth and also with soil drying
427 (Bengough *et al.*, 2006; Lynch and Wojciechowski, 2015). The ability of roots to penetrate
428 hard soils is therefore an important aspect of water capture under drought, especially
429 water capture from deep soil strata. Phenotypes like RCA and RCS that reduce cortical burden
430 in mature root segments are not likely to affect soil penetration, which is a function of the
431 root tip. Anatomical phenotype states like reduced CCFN could reduce root diameter, which
432 would be expected to reduce penetration of hard soil (Bengough *et al.*, 2006). In this
433 context, it is noteworthy that several anatomical phenotypes were related to the biomechanical
434 properties of maize roots and their ability to penetrate hard wax layers independently of
435 root diameter, including cell size in the distal cortex, cortical cell wall area, and stele
436 diameter (Chimungu *et al.*, 2015a). The architectural phenotype states of reduced axial root
437 and lateral root production may enable remaining roots to have larger diameter, which
438 would improve their penetration of hard soil. Phenotype states that reduce radial water
439 transport, like RCA and RCS, may permit growing root tips to remain hydrated and would
440 allow them to penetrate soil with greater water content and therefore less impedance
441 (Lynch *et al.*, 2014).

442 **2f. Root phenotypes that optimize water capture may also optimize N capture**

443 Suboptimal N availability is second only to water as a global limitation to crop productivity.
444 In low-input systems, suboptimal N limits yield, while in high-input systems, intensive N
445 fertilization is a primary economic, energy, and environmental cost of the production of
446 non-leguminous crops (Hirel *et al.*, 2011; Cassman *et al.*, 2002; Ribaud *et al.*, 2011). In
447 most agroecosystems N accumulates in deeper soil strata over time as a result of
448 leaching (Thorup-Kristensen and Kirkegaard 2016), which led to the proposal that deeper
449 rooting will improve the capture of both N and water (Lynch 2013). For example, steeper
450 root growth angles improve water capture in several crop species (*e.g.*, Manschadi *et al.*,
451 2006; Singh *et al.*, 2012), and in maize also improve N capture (Trachsel *et al.*, 2013;
452 Dathe *et al.*, 2016). In maize, root phenes that regulate root depth and thereby the capture
453 of both water and N when those resources are limiting include the number of nodal roots
454 (Gao and Lynch 2016; Saengwilai *et al.*, 2014b), lateral root branching density (Zhan *et*
455 *al.*, 2015; Zhan and Lynch 2015), and root cortical aerenchyma (Zhu *et al.*, 2010;
456 Saengwilai *et al.*, 2014a; Chimungu *et al.*, 2015b). Optimization of root phenotypes for
457 water capture may therefore also improve N capture, with benefits for both high-input and
458 low-input agroecosystems.

459 **3. The challenges of low-input systems**

460 Many important agroecosystems do not receive intensive inputs. These include systems
461 in which intensive management is possible yet is uneconomical, such as pastures,
462 rangeland, or production of biofuel crops, and systems in which intensive use of inputs is
463 not an option because of lack of capital, lack of expertise, or poor availability of inputs, as
464 occurs in smallholder agriculture in developing nations. Such systems suffer from many
465 of the constraints found in natural ecosystems, including multiple biotic and abiotic
466 stresses, intense weed competition, and spatiotemporal variability in the availability of soil
467 resources, especially water. As a result, optimal root phenotypes for drought resistance
468 in high-input systems may be suboptimal in low-input systems.

469 Phosphorus availability is a key difference between high-input and low-input systems.
470 Intensive P fertilization is characteristic of high-input agriculture, whereas low P
471 availability is a primary constraint to crop productivity in the majority of low-input systems.

472 This is because of the inherently low P availability in the weathered soils characteristic of
473 the tropics and subtropics, in addition to ongoing soil degradation. Root strategies to
474 acquire P have several important tradeoffs for water acquisition. Phosphorus is an
475 immobile soil resource that concentrates in the topsoil over time, whereas water is highly
476 mobile and moves to deep soil strata over time. The low mobility of P in soil means that
477 roots (and their symbionts) must be in close proximity to P in order to acquire it (Barber,
478 1995), while water can move to the root from much larger distances, thereby creating
479 tradeoffs for intensive vs. extensive soil foraging strategies. A third tradeoff is that root
480 uptake creates P depletion zones that only slowly replenish, whereas soil domains
481 depleted of soil water may be rapidly replenished. This means that continued root growth
482 and exploration of undepleted soil domains is more important for P capture than for water
483 capture.

484 Clear tradeoffs for water and P capture are evident for several root architectural phenes.
485 Shallow root growth angles promote topsoil foraging and P capture (Lynch, 2011),
486 whereas steep root growth angles promote subsoil foraging and water capture (Ho *et al.*,
487 2005). In maize, dense lateral root branching promotes P capture (Postma *et al.*, 2014),
488 whereas sparse lateral branching promotes water capture (Zhan *et al.*, 2015). Production
489 of few axial roots is beneficial for water capture by allowing root axes to grow to greater
490 soil depths (Gao and Lynch, 2016), whereas increased nodal root number improves P
491 capture (Bayuelo-Jiménez *et al.*, 2011). Anatomical phenes that reduce cortical
492 parenchyma, like RCA, RCS, and reduced CCFN, are beneficial for water capture but
493 may reduce symbiotic P capture by reducing mycorrhizal habitat. In contrast, anatomical
494 phenes that reduce the metabolic cost of soil exploration should have benefits for the
495 capture of both mobile and immobile resources. For example, RCA is beneficial for the
496 capture of the mobile resources water (Zhu *et al.*, 2010; Chimungu *et al.*, 2015b) and N
497 (Saengwilai *et al.*, 2014a), while also being beneficial for the capture of the immobile
498 resources P and K (Postma and Lynch, 2010, 2011). Root hairs are useful for capture of
499 P (Bates and Lynch, 2000a; Miguel *et al.*, 2015) as well as water (Carminati *et al.*, 2017),
500 while incurring little direct metabolic cost (Bates and Lynch, 2000b).

501 In considering the optimal root phenotype for low-input systems it must be considered
502 that many such systems employ polycultures composed of different crop species and/or
503 different genotypes of the same species instead of the genetic monocultures
504 characteristic of high-input systems. The fitness value of root phenotypes in polycultures
505 may differ from monocultures due to resource competition and complementarity, and
506 fitness impacts upon the polyculture as a whole rather than just to one species. For
507 example, the ancient maize/bean/squash polyculture is more efficient in capturing mineral
508 N than are the component single species because of niche complementarity (Postma and
509 Lynch, 2012; Zhang *et al.*, 2014). It is likely that water capture by polycultures is also
510 affected by interactions among the roots of each species. In addition to niche
511 complementarity it is possible that members with deeper and shallower roots may benefit
512 each other through root-mediated hydraulic redistribution (Brooker *et al.*, 2015).

513 The greater intensity of biotic and abiotic stress in low-input systems, and greater
514 variability in the distribution of soil resources in time and space, may make the production
515 of numerous axial roots and plasticity in response to local resource availability more
516 important than they are in high-input systems. Better understanding of the fitness
517 landscapes of root phenotypes in the progression of wild taxa in natural systems through
518 landraces in low-input systems to elite monogenetic stands in high-input systems would
519 be useful in resolving these issues, and in guiding the development of more drought
520 resistant crops in developing nations.

521 **4. Future Prospects**

522 The need for drought resistant crops is critical, and will surely grow in years ahead
523 because of a growing human population with increasing food demand per capita, ongoing
524 degradation of soil and water resources, and the accelerating effects of global climate
525 change. Crops with improved root phenotypes will be an important element of future
526 agroecosystems. In recent years significant progress has been achieved in discovering
527 specific root phenes, phene states, and integrated phenotypes for superior water capture.
528 Ideotype breeding, grounded in an understanding of the fitness landscape of specific root
529 phenes in specific phenotypic, environmental, and management contexts, would be an

530 effective path to deploy root phenotypes in crop breeding (Cooper et al. 2014; Lynch,
531 2015). For this to succeed it is important that phenes, elemental units of the phenotype,
532 are evaluated rather than phene aggregates, which are under more complex genetic
533 control and have more complex environmental interactions. For example, although root
534 depth is generally viewed as a 'secondary trait' that offers mechanistic insight into drought
535 resistance, recent research shows that root depth is in fact regulated by multiple
536 independent phenes and phenes states (Lynch and Wojcieszowski, 2015), whose
537 interactions with each other and with the environment are quite complex (Thorup-
538 Kristensen and Kirkegaard, 2016). Better definition of target phenes would greatly
539 facilitate both genetic and functional analysis. An excellent example of this approach is
540 the work at IRRRI to understand how specific QTL affect resistance to specific types of
541 drought in specific production environments by regulating specific root phene states such
542 as root growth angle and nodal root number (e.g. Catolos et al 2017).

543 In this article, I propose that the optimal root phenotype for drought resistance in high-
544 input agroecosystems is fundamentally different from ancestral root phenotypes that have
545 been selected in natural ecosystems and traditional low-input agroecosystems. A
546 possible exception is wild taxa adapted to arid environments. Many arid environments are
547 characterized by severe water limitation but good availability of other soil resources and
548 relatively low plant density, hence relatively slight belowground competition, and less soil
549 biotic activity, hence reduced root loss. Therefore, they may possess the parsimonious
550 root phenotype that I propose here. Several important crop species have wild relatives
551 adapted to arid conditions. It would be useful to study the root phenotypes of such taxa,
552 although their deployment in elite crop breeding could be complex. In some cases,
553 cultivated taxa exist that are related to principal crops, that might serve as models or
554 genetic resources for crop breeding. For example, the tepary bean, *Phaseolus acutifolius*,
555 is a close relative of the common bean, *Phaseolus vulgaris*, but is native to semi-arid
556 regions of North America, and has much greater resistance to heat and drought than does
557 common bean (Pratt and Nabhan, 1988; Federici *et al.*, 1990; Rao *et al.*, 2013).
558 Interspecific hybridization of tepary bean with common bean has led to the development
559 of common bean lines with substantially improved resistance to drought and heat (Mejia-
560 Jimenez *et al.*, 1994; Muñoz *et al.*, 2004; Rao *et al.*, 2013). Other cases exist in which

561 principal global crops are genetically related to other crop taxa from semiarid regions, as
562 is the case with maize (*Zea mays*) vs. sorghum (*Sorghum bicolor*). Comparative analysis
563 of root phenotypes from such taxa would be useful in identifying phenes and potentially
564 genes for improvement of the drought resistance of a target species. In addition to
565 analysis of related taxa from semiarid environments, it would be useful to profile root
566 phenotypes of a diverse set of elite lines and landraces within crops, to establish the
567 range of variation present and identify patterns associated with drought resistance. This
568 has not often been attempted, possibly because of the difficulty in phenotyping roots
569 under realistic conditions. The advent of high-throughput methods to quantify root phene
570 states under realistic conditions is a promising development in this context (e.g. Bucksch
571 *et al.*, 2014).

572 The use of modern *in silico* approaches will be invaluable in testing the validity of the
573 ideas proposed here, and more generally in understanding the fitness landscape of root
574 phenotypes. This is because of the large number of interactions among specific root
575 phenes and the environment, and the large number of environmental scenarios of
576 interest, including different soil water regimes as well as future environments with drought
577 coinciding with elevated atmospheric CO₂ and high heat, both of which could strongly
578 interact with drought stress, as well as novel management scenarios. Modeling is
579 particularly attractive for drought, because of the difficulty of imposing managed drought
580 regimes in the field and in replicating realistic drought stress in controlled environments.
581 To be useful in this context, models should faithfully capture the essential elements of the
582 acquisition and transport of water by roots in drying soil, shoot responses to water stress,
583 as well as interactions of roots with shoots. Presently no models exist with these
584 capabilities at the level of detail that would enable evaluation of specific phenes and
585 phene states, or that could faithfully model emergent properties resulting from soil-root-
586 shoot interactions (Tardieu *et al.*, 2017). Parsimony is a valuable quality in heuristic
587 models, because of the difficulty in understanding and interpreting results from models
588 with many variables, as well as the problem of error propagation. To paraphrase
589 Einstein's famous quote regarding theories, 'a model should be as simple as possible,
590 but no simpler'. Rather than construct complex models attempting to simulate all relevant
591 processes, a more feasible and robust option would be to integrate a family of models,

592 each focusing on a distinct set of processes, within a larger framework capable of
593 exchanging data among the submodels, checking for error propagation, and capturing
594 emergent properties. Increasingly robust models exist for water movement in soil, canopy
595 responses to water stress, and the capture of soil resources by roots. The rapid
596 development of functional-structural plant models is a very promising development in this
597 context (e.g. (Dunbabin *et al.*, 2013; Marshall-Colon *et al.*, 2017; Postma *et al.*, 2017).

598 I propose that a parsimonious root phenotype would be advantageous for drought
599 resistance in high-input agroecosystems. These concepts may also apply to the shoot
600 phenotype. Reduced shoot branching, reduced leaf production, greater turnover of
601 older/shaded leaves, reduced investment in parenchyma tissue in the stem and in leaves,
602 narrower leaves that permit better light penetration of the canopy while improving leaf
603 energy budgets under drought, reduced stem production and elongation in species that
604 are not already dwarfed, *etc.*, could result in greater drought resistance without
605 compromising yield potential by focusing resource allocation to the most productive
606 tissues. For example, reduced tillering and leaf size improves drought resistance in
607 sorghum by reducing soil water depletion prior to anthesis (Borrell *et al.*, 2014). As with
608 parsimonious root phenotypes, parsimonious shoot phenotypes may not be
609 advantageous in low-input agroecosystems however.

610 **5. Conclusion/summary**

611 I propose that parsimonious root phenotypes would be advantageous for drought
612 resistance in high-input agroecosystems. Selection regimes for crop ancestors and
613 landraces include an array of biotic and abiotic stresses, intense belowground
614 competition, and spatiotemporal variability in the distribution of soil resources. These
615 factors favored phenotypes with prolific production of axial and lateral roots,
616 developmental plasticity in response to local resource availability, and maintenance of
617 unspecialized root tissues such as cortical parenchyma. Phosphorus in particular has
618 important tradeoffs with water for root structure and function, since water is highly mobile,
619 whereas phosphorus is highly immobile. High-input agroecosystems have removed many
620 of these constraints to root function. Intensive fertilization removes P limitation and other

621 abiotic limitations to root function like soil acidity. Pesticide application reduces root loss
622 to herbivores and pathogens, and belowground competition from weeds. High-density
623 monocultures insure that soil resources lost to neighboring plants still contribute to crop
624 production. Management regimes create regular and predictable resource availability in
625 time and space. However, drought stress remains a primary risk to crop production in
626 rainfed agriculture, and this risk is likely to grow in the future. Therefore, root phenotypes
627 that focus on water capture at the expense of ancestral adaptations would be better suited
628 to high-input agroecosystems. Parsimonious architectural and anatomical phenotypes
629 would permit greater resource allocation to deeper roots, which in most agroecosystems
630 results in greater water capture. Specific phenotypes contributing to parsimonious
631 architectural phenotypes include fewer axial roots, reduced density of lateral roots,
632 reduced growth responsiveness to local resource availability, and greater loss of roots
633 that do not contribute to water capture, for example in dry topsoil. Specific phenotypes
634 contributing to parsimonious anatomical phenotypes include reduced production of
635 cortical parenchyma through reduced production of cortical cell files, greater loss of
636 cortical parenchyma through formation of root cortical aerenchyma and root cortical
637 senescence, and larger cortical cell size. Parsimonious root phenotypes may be less
638 useful in low-input agroecosystems, which are characterized by multiple challenges and
639 tradeoffs for root function in addition to water capture. Although some of these ideas are
640 supported by empirical evidence, they remain largely hypothetical. Analysis of the fitness
641 landscape of specific root phenotypes, phenotypes and integrated phenotypes is a complex
642 research challenge that will be aided by the development of robust functional-structural
643 models capable of simulating the dynamics of root-soil interactions.

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Figure Legends

Figure 1. Visualization of maize root phenotypes 42 days after germination in *OpenSimRoot*. Phenotype (a) has 25 crown roots, 10 lateral roots per cm of axial root, and 1 cm distance from stem to first lateral emergence. Phenotype (b) has 10 crown roots, 5 lateral roots per cm of axial root, and 0.5 cm distance from stem to first lateral emergence. Images are the same scale. These phenotypic values have been observed in plants in the field (see citations in text). Plants were simulated at 105 kg ha⁻¹ soil N availability, which represents moderate N stress, to illustrate the effect of these phenotypic states on root development when soil resources are limited. Image courtesy of Xiyu Yang.

Figure 2. Correlations between crown root number and shoot biomass of maize at 35 days after planting in greenhouse mesocosms (A), and shoot biomass (B) and relative yield (WS:WW, %) (C) at anthesis in the field under water-stressed conditions. Each point is means of four replicates of each genotype \pm SE. From Gao and Lynch 2016.

Figure 3. Visualization of maize root phenotypes 42 days after germination in *SimRoot*, showing the effects of varying the number of crown roots from 46 (left) to 6 (right) on root depth and lateral root development. Roots are shown reflecting back to the center to represent roots from neighboring plants of the same phenotype. Hotter colors represent more N capture. Image courtesy of Larry York.

Figure 4. Nodal root number has declined over the past 100 years of maize breeding. Dotted lines represent planting density with gaps between dots being proportional to differences in densities, with 20K, 40K, and 80K being 20,000, 40,000, and 80,000 plants ha⁻¹. Triangles are in low nitrogen and circles in high nitrogen. Points represent the mean of the four varieties in that Era time period in the specific nitrogen and density combination, and vertical lines the standard error. Letters demonstrate a significant difference between the first and last Era periods based on a *t*-test ($p = 0.01541$) conducted after ANOVA demonstrated a significant effect of Era period. Presence or absence of an asterisk next to a treatment in the legend indicates whether a treatment effect is significant or not, respectively. From York *et al.* 2015.

Figure 5. Effects of specific root phenes on shoot mass of modern maize cultivars and cultivars from 100 years ago as simulated in *SimRoot*. Plants were simulated in a current environment of very high density (120,000 plants ha⁻¹) and high nitrogen (120K HN), and a historic environment of low density (40,000 plants ha⁻¹) and low nitrogen (40K LN). Solid bars represent the means with lines being the standard error. Abbreviations at the bottom of the bar give the phenotype, as follows. The *Old* (O) phenotype has a steeper angle and a few more nodal roots than the *Modern* (M) phenotype. *Old* and *Modern* have low aerenchyma. *Old* + Angle (OA) is the same as the *Old* phenotype but with the same shallow angle as the *Modern*, while *Old* + NRN (ON) is the same as the *Old* phenotype but with the same fewer nodal roots as the *Modern*. *Modern* + RCA (MR) is the *Modern* phenotype but with high aerenchyma. From York *et al.* 2015.

Figure 6. Correlation between lateral root branching density of maize crown roots and A: relative shoot dry weight (% relative to greatest shoot dry weight within each location) in greenhouse mesocosms (GH), in the field in Arizona (AZ) and Pennsylvania (PA), B: relative yield (% relative to greatest yield) in PA under water stress conditions. Each point is the mean of four replicates of each genotype. From Zhan, Schneider, Lynch 2015.

Figure 7. Phenotypic variation in maize for Root Cortical Aerenchyma (RCA, 7a), Cortical Cell File Number (CCFN, 7b) and Cortical Cell Size (CCS, 7c). 7d: Under water stress genotypes with greater RCA have less respiration (nmol CO₂ s⁻¹ cm⁻¹), deeper rooting (7a: cm roots at 40-50 cm soil depth; 7b,c: D₉₅, which is the depth in cm attained by the 95th percentile of roots), and greater yield (g/plant), as did genotypes with reduced CCFN (7e) and greater CCS (7f). Data shown are means ± SE (n= 3 or 4). Means with different letters are significantly different (p ≤0.05). Redrawn from Zhu, Brown, Lynch 2010 and Lynch 2015.

Figure 8. Correlation between maize yield and root cortical aerenchyma (% of cortical cross-sectional area) under water stress condition in two field environments in Malawi: **(A)** Bunda and **(B)** Chitala. From Chimungu *et al.* 2015b.

Figure 9. Root cortical senescence in barley reduces root respiration and the radial transport of water and nutrients, yet permits greater root depth and nutrient capture. From Schneider and Lynch, in press.